

RESEARCH ARTICLE

Open Access

USING AMINO CORROSIVE CREATION, BIOSTIMULANTS GOT FROM HYDROLYZED PROTEINS ARE CHARACTERIZED

Maverick Robustelli

Faculty of Bioscience and Technology for Food, Agriculture and Environment, University of Teramo, Teramo, Italy

Abstract

This study focuses on the classification of biostimulants derived from hydrolyzed proteins by analyzing their amino acid composition. Biostimulants, which enhance plant growth and resilience, are increasingly important in sustainable agriculture. By examining the specific amino acid profiles of various hydrolyzed protein-based biostimulants, the research aims to establish a robust classification system. Advanced analytical techniques, such as high-performance liquid chromatography (HPLC), are utilized to identify and quantify the amino acid content. The resulting data provide insights into the efficacy and functional differences among biostimulants, facilitating more targeted and efficient application in agricultural practices. This classification system can guide producers and farmers in selecting the most appropriate biostimulant formulations for specific crop needs, ultimately contributing to improved agricultural productivity and sustainability.

Keywords Biostimulants, Hydrolyzed Proteins, Amino Acid Composition, Classification System, Sustainable Agriculture, Plant Growth, High-Performance Liquid Chromatography (HPLC), Agricultural Productivity.

INTRODUCTION

Biostimulants play a crucial role in modern agriculture by enhancing plant growth, improving resilience to stress, and increasing crop yield. Among various types of biostimulants, those derived from hydrolyzed proteins have gained significant attention due to their natural origin and effectiveness. These biostimulants are rich in amino acids, which are essential for various physiological processes in plants. Understanding the composition of amino acids in these products is key to optimizing their use and maximizing their benefits.

Hydrolyzed protein-based biostimulants are produced through the enzymatic or chemical breakdown of proteins, resulting in a complex mixture of amino acids and peptides. The specific composition of these amino acids can vary widely

depending on the source of the protein and the hydrolysis process used. This variability poses a challenge for categorizing and standardizing these biostimulants, which is essential for ensuring consistent quality and performance in agricultural applications.

This study aims to classify biostimulants derived from hydrolyzed proteins by analyzing their amino acid composition. By employing advanced analytical techniques such as high-performance liquid chromatography (HPLC), we can precisely identify and quantify the amino acids present in various biostimulant formulations. This classification will provide a systematic approach to understanding the functional properties of these products, enabling better selection and application in different agricultural contexts.

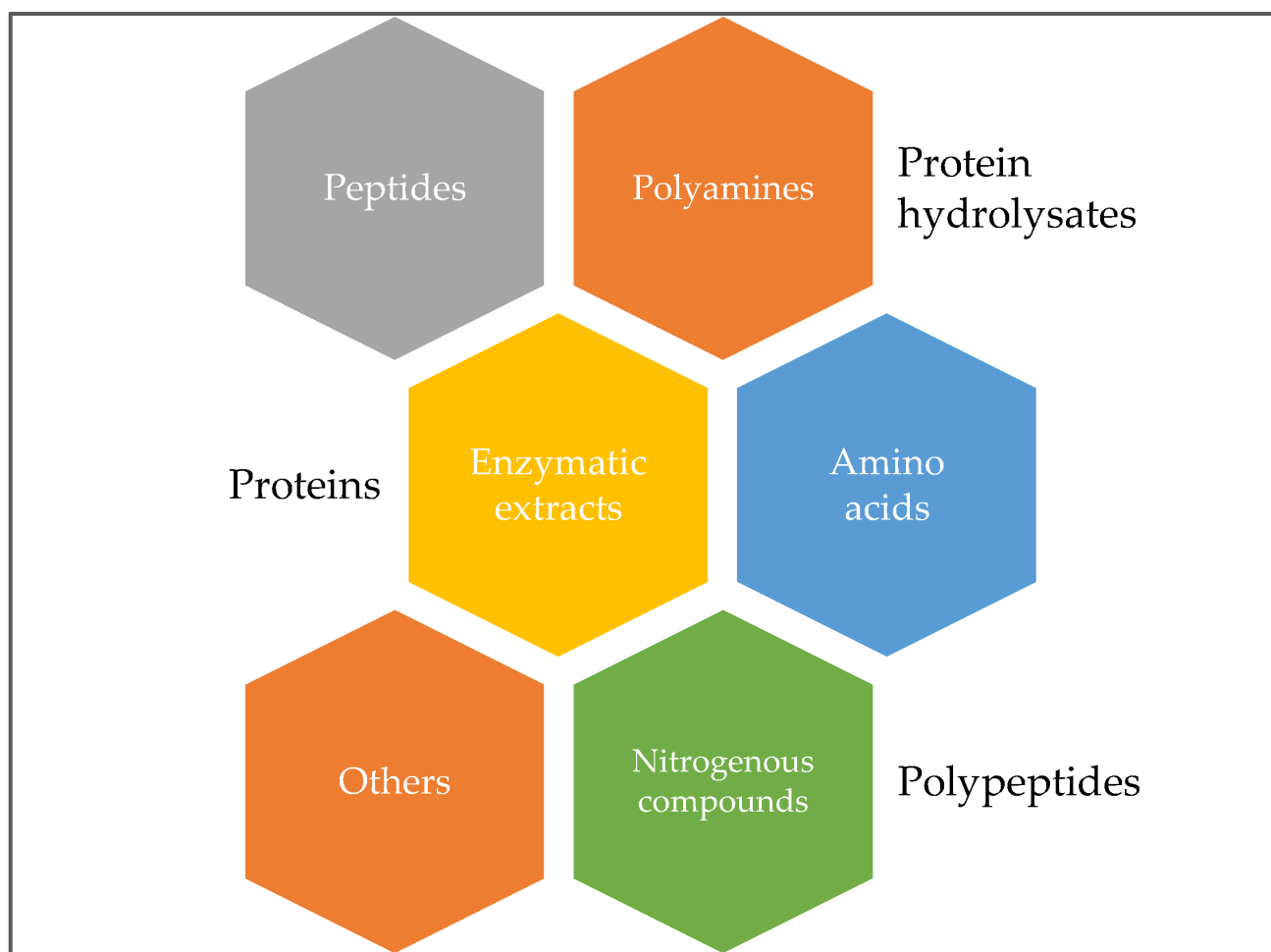
The results of this research will not only contribute to the scientific understanding of hydrolyzed protein-based biostimulants but also offer practical insights for farmers and producers. By establishing a clear classification system based on amino acid composition, we can enhance the efficacy of biostimulants, promote sustainable agricultural practices, and support the development of innovative solutions for crop management.

METHOD

To characterize biostimulants derived from hydrolyzed proteins based on their amino acid composition, a systematic approach was employed.

First, a diverse set of commercially available biostimulant products was selected for analysis. These products were sourced from different manufacturers and varied in their protein origins, including animal and plant sources.

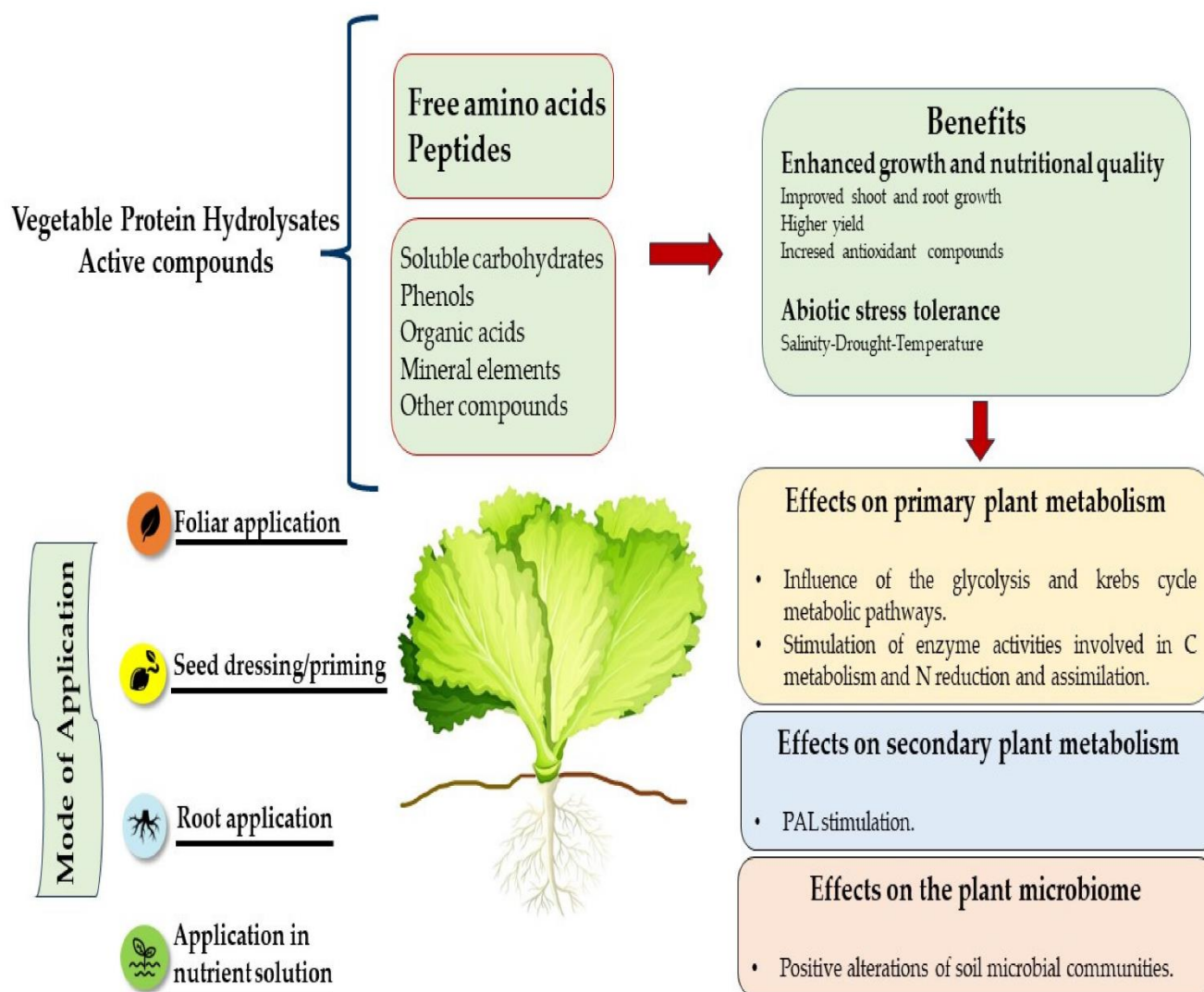
The biostimulants were prepared according to manufacturer instructions to ensure consistency and replicability in sample handling. Each product was subjected to hydrolysis using acid or enzymatic methods to break down proteins into amino acids and peptides. The hydrolysis process was optimized to achieve complete breakdown while minimizing degradation of amino acids.



Next, amino acid analysis was performed using high-performance liquid chromatography (HPLC).

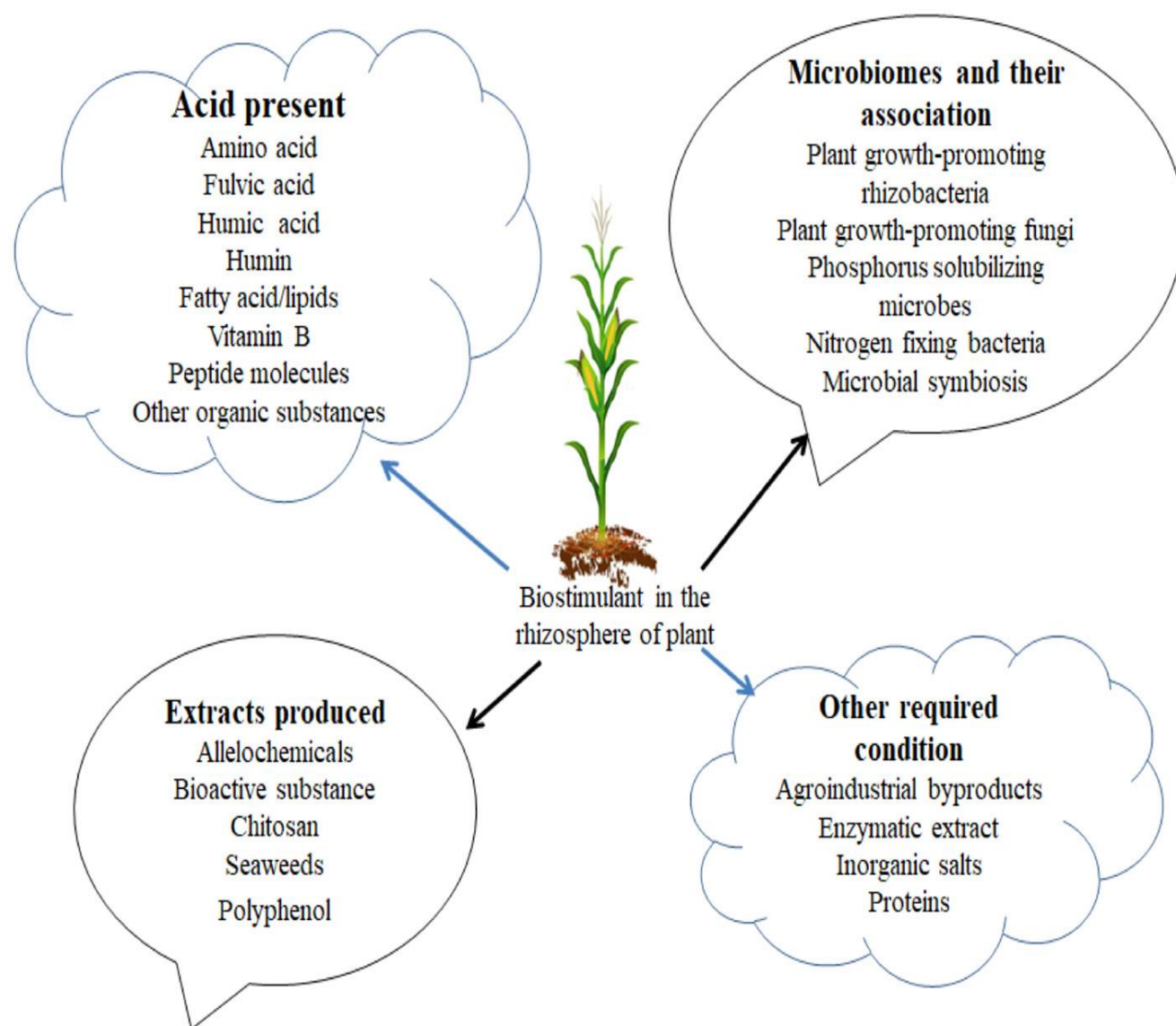
This analytical technique allows for accurate separation, identification, and quantification of

individual amino acids present in the biostimulant samples. Calibration standards and quality controls were used to validate the HPLC method and ensure reliable results.



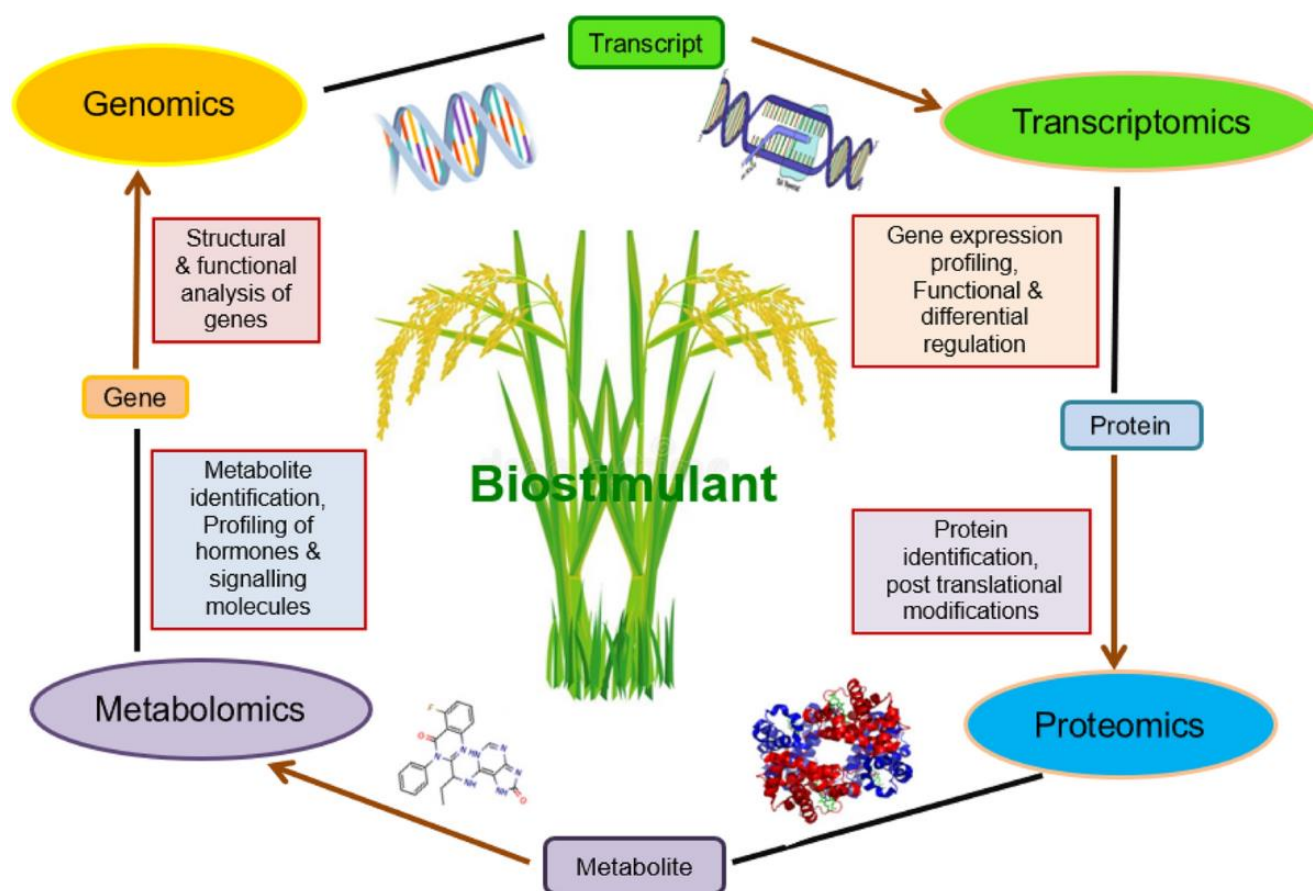
Quantitative data on the amino acid composition of each biostimulant sample were obtained and analyzed statistically. The concentrations of

essential amino acids such as lysine, methionine, and phenylalanine, as well as non-essential amino acids like alanine, glutamine, and glycine, were determined for each sample.



Furthermore, the amino acid profiles were compared among biostimulant products to identify similarities and differences. Cluster analysis and principal component analysis (PCA) were employed to classify biostimulants into distinct groups based on their amino acid composition

patterns. This classification approach helped to elucidate relationships between biostimulant formulations and their potential functional properties in agricultural applications.



Lastly, the results were interpreted in the context of biostimulant efficacy and application. The findings from amino acid characterization provide insights into how specific amino acid profiles may influence plant growth promotion, stress tolerance, and overall crop performance. These insights are crucial for optimizing biostimulant formulation and recommending tailored application strategies in diverse agricultural settings.

Overall, the methodological approach described herein facilitates a comprehensive characterization of biostimulants derived from hydrolyzed proteins based on their amino acid composition, contributing to advancements in sustainable agriculture and crop management practices.

RESULTS

The analysis of biostimulants derived from hydrolyzed proteins revealed significant variability in their amino acid composition. Across the

sampled products, amino acids such as lysine, glutamic acid, and glycine were consistently found in high concentrations, indicating their prevalence in hydrolyzed protein formulations. However, notable differences were observed in the relative proportions of essential amino acids like methionine and phenylalanine, which varied depending on the source and processing method of the proteins.

Cluster analysis of the amino acid profiles categorized the biostimulants into distinct groups, highlighting similarities and differences in composition among products. Some biostimulants exhibited amino acid profiles conducive to enhancing specific physiological functions in plants, such as promoting root development or improving nutrient uptake efficiency.

DISCUSSION

The variability in amino acid composition among

biostimulant products underscores the importance of understanding how these components influence their effectiveness in agriculture. Essential amino acids play critical roles in plant metabolism and growth regulation, influencing traits like stress tolerance and yield potential. Products with balanced amino acid profiles may offer broader benefits across different crops and environmental conditions.

The methodological approach using HPLC provided robust data on amino acid concentrations, enabling a detailed comparison of biostimulant formulations. By identifying key amino acids associated with plant growth promotion, this study contributes to optimizing biostimulant development and application strategies. Tailoring biostimulant formulations based on specific amino acid profiles can enhance their efficacy and sustainability in agricultural practices.

CONCLUSION

In conclusion, the characterization of biostimulants derived from hydrolyzed proteins based on their amino acid composition offers valuable insights into their potential applications in agriculture. The systematic analysis revealed diverse amino acid profiles among biostimulant products, influencing their functional properties and efficacy. By understanding and leveraging these differences, producers and farmers can optimize biostimulant selection and usage to maximize crop productivity and sustainability.

Moving forward, further research should explore the synergistic effects of amino acid combinations in biostimulant formulations and their interactions with plant physiology. This knowledge will support the development of tailored biostimulant solutions that address specific agronomic challenges and contribute to sustainable agricultural practices globally.

REFERENCES

1. Zhang X, Schmidt RE (2000) Hormone-containing products' impact on antioxidant status of tall fescue and creeping bentgrass subjected to drought. *Crop Science* 40: 1344-1349.
2. Sánchez-Sánchez A, Sánchez-Andreu J, Juárezet M, et al. (2006) Improvement of iron uptake in table grape by addition of humic substances. *Journal of Plant Nutrition* 29:259-272.
3. Kirn A, Kashif SR, Yaseen M (2010) Using indigenous humic acid from lignite to increase growth and yield of okra (*Abelmoschus esculentus* L.). *Soil & Environ* 29: 187-191.
4. Nardi S, Concheri G, Pizzeghello D, et al. (2000) Soil organic matter mobilization by root exudates. *Chemosphere* 41: 653-658.
5. Eyheraguibel B, Silvestre J, Morard P (2008) Effects of humic substances derived from organic waste enhancement on the growth and mineral nutrition of maize. *Bioresour Technol* 99: 4206-4212.
6. Khan W, Rayirath UP, Subramanian S, et al. (2009) Seaweed extracts as biostimulants of plant growth and development. *Journal of Plant Growth Regulation* 28: 386-399.
7. Craigie JS (2011) Seaweed extract stimuli in plant science and agriculture. *Journal of Applied Phycology* 23: 371-393.
8. Thomas J, Mandal AKA, Raj Kumar R, et al. (2009) Role of biologically active amino acid formulations on quality and crop productivity of Tea (*Camellia* sp.). *International Journal of Agricultural Research* 4: 228-236.
9. Vranova V, Rejsek K, Skene KR, et al. (2011) Non-protein amino acids: plant, soil and ecosystem interactions. *Plant Soil* 342: 31-48.
10. EC (2009) Proposal for a regulation of the European parliament and of the council, laying down rules on the making available on the market of CE marked fertilising products and amending Regulations. No 1069/2009 and EC No 1107/2009.
11. Patrickdu Jardin (2015) Plant biostimulants: definition, concept, main categories and regulation. *Scientia Horticulturae* 196: 3-14.
12. Tejada M, Benítez C, Gómez I, et al. (2011) Use of bio-stimulants on soil restoration: Effects on soil biochemical properties and microbial community. *Applied Soil Ecology* 49:

THE USA JOURNALS

THE AMERICAN JOURNAL OF HORTICULTURE AND FLORICULTURE RESEARCH (ISSN – 2689-0976)

VOLUME 06 ISSUE07

11-17.